

Systems Engineering

**Computational Visualization Method for Drug Discovery**

Final Paper

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*“Fortune Favors the prepared mind.”*

Louis Pasteur

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# Abstract

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**Computational visualization method for drug discovery**

by Shy Alon

The process of drug design keeps growing in difficulty and length and the average cost of a new successful drug is estimated by the pharmaceutical companies to be 5.5 billion USD. That high cost includes the cost of testing thousands and sometimes millions of compounds that may be screened and assessed early in the R&D process, only a few of which will ultimately receive approval.

Of the four steps of drug development process (discovery, pre-clinical research, clinical research and FDA review) the discovery phase is considered by the industry to be so risky and unprofitable that the pharmaceutical industry has abandoned it completely to non-government organizations (NGOs) and academic institutions which pursue the discovery of new drugs for motives other than profit.

The hypothesis of this work is that a cost effective computational method could be utilized to significantly reduce the costs of the pre-clinical phase to the degree they would be cost effective. The method we used is a classification algorithm utilizing a combination of a feature selection mechanism (a genetic algorithm) with an optimized dimensionality reduction algorithm (t-SNE) and a nearest neighbor classifier.

Having used a database of over seven thousand compounds (specifically picked for their interaction with GPCR compounds) with over one hundred and thirty features we have shown that a computational method can be used to effectively predict whether a specific compound will have a certain effect with a level of certainty above ninety percent. By automatically selecting the relevant features for each target GPCR we demonstrated that in the 2-dimensional space (after using dimensionality reduction with t-SNE) the nearest neighbors were very likely to share the same effect.

# Introduction

## Motivation

The cost of a new drug is estimated at 5.5 billion USD (Winegarden, 2014) and only two out of ten new drugs cover their incurred R&D expenses. According to the U.S. food and drug administration (FDA) before a new drug hits the market there are 4 required steps:

1. Discovery and development: Discovery of new drugs. At this stage in the process, many thousands of compounds are potentially candidates for development as a medical treatment. Early testing filters the candidates to a small number of compounds. Once a promising compound is identified experiments are conducted to gather information on how it is absorbed, distributed, metabolized, and excreted; Its potential benefits; the best dosage; the best way to give the drug (such as by mouth or injection); side effects; effects on different groups of people (such as by gender, race, or ethnicity); interaction with other drugs and treatments and its effectiveness as compared with similar drugs.
2. Preclinical Research: Verify compound’s toxicity
3. Clinical Research: The studies, or trials, that are conducted on people.
4. FDA Review: Application to market the drug.

The discovery phase costs, as reported by pharma market members, as high as one billion US dollars (English, Lebovitz, Griffin, 2010) and is a liability that the pharmaceutical companies cannot accept until its effectiveness is dramatically.It is safe to assume that every test for a candidate compound costs roughly 10,000 USD and right now it takes a corpus of 125,000 candidates to produce 75 viable pre-clinical trials candidates (Winegarden, 2014). Out of these one is likely make it to production. This means that under present conditions finding a sufficiently large viable candidate group would cost 1.25 billion U.S dollars.

In order to make the discovery phase economically viable we seek to reduce the cost by one order of magnitude – which means automatically and cheaply filter most of the candidates and leave at most one tenth of the candidate population – without losing viable candidate.

## Research Hypothesis

The proposed research will address a clear need for new computational cost effective method for the drug discovery stage. Such a method will be able to predict the biological activity of a compounds prior to their synthesis. Visualization of chemical data and a good representation of the chemical space is useful in many drug design applications including the selection of compounds for synthesis, the selection of compounds for biological evaluation and selection of subsets for the design of information-rich compound libraries. Here we present a new computational method, namely t-Distributed Stochastic Neighbor Embedding (t-SNE) optimization method for the visualization and the representation of the chemical space for the selection of compounds for the drug discovery stage. In order to get a good representation of the chemical space we have coupled the t-SNE algorithm with an optimization engine for feature selection.

## Research Objectives

In order to meet the global goals of the current research, the following specific goals were defined:

1. Development of a new optimization-based method for visualization of big data, namely: t-Distributed Stochastic Neighbor Embedding (t-SNE) optimization method.
2. Evaluation of the new t-Distributed Stochastic Neighbor Embedding (t-SNE) optimization method for its ability to reduce dimension, to cluster and to classify the data.
3. Application of the new method for the analysis of different G-protein-coupled receptors (GPCRs) families.

## Application

In order to test our hypothesis a. G-protein-coupled receptors (GPCRs) datasets were selected. As will be discussed below in the literature review section, GPCRs are the largest single superfamily of proteins. Over 1000 GPCRs have been identified in vertebrates, in which they regulate a wide variety of signals including ions, amino acids, nucleotides, peptides, lipids, hormones, neurotransmitters, and chemokines (Bockaert, Pin, 1999). It has been estimated that GPCRs, while being only around 3% of known molecular targets, represent as much as 45% of current drug targets (Hopkins, Groom, 2002; Drews, 2000). Due to their excellent potential for drug discovery, GPCR targets represent up to 30% of the portfolio of many pharmaceutical companies (Klabunde, Hessler, 2002).

The research hypothesis discussed in this paper, in the context of GPCRs activation focused pharmaceutical industry research, is that it is possible to optimize a feature selection for a dimensionality reduction algorithm so that the compounds selected according to the distance from tagged compounds present the best candidate for pre-clinical trials. This is accomplished by building an algorithmic candidate and testing it on a rich enough database.

# Literature review

## Pharmaceutical Research Challenge

According to the U.S. food and drug administration (FDA) before a new drug hits the market there are 4 required steps (see figure 1):

Figure Pharma Compound Funnel

1. Discovery and development: discovery of new drugs through new insights into a disease process that allow researchers to design a product to stop or reverse the effects of the disease, broad range of tests of molecular compounds to find possible beneficial effects for multiple diseases or existing treatments that have unanticipated effects or new technologies.
2. Preclinical Research: Before human trials the compound’s toxicity must be ascertained using two types of preclinical research: in vitro (using controlled environment outside of a living organism) and in vivo (using living organisms such as cells and animals). Preclinical studies are limited in scope but must provide detailed information on dosing and toxicity levels, leading to the decision whether the drug should be tested in people.
3. Clinical Research: The studies, or trials, that are conducted on people. Clinical trials vary greatly on scales of risk and process and follow a typical series from early, small-scale, Phase 1 studies to late-stage, large scale, Phase 3 studies.
4. FDA Review: when a drug has indicated from its early tests and preclinical and clinical research that it is safe and effective for its intended use, the developer can file an application to market it.

The challenges facing pharmaceutical research were discussed in many papers published by PhRMA, an institution representing many American pharma companies. In their industry profile for 2017 PhRMA address the “The Lengthly, Costly and Uncertain Biopharmaceutical Research and Development Process” and its negative impact on the industry.

According to PhRMA the process for researching and developing new medicines keeps growing in difficulty and length. On average, it takes at least ten years for a new medicine to complete the journey from initial discovery to the marketplace, with clinical trials alone taking six to seven years on average. The average cost to research and develop each successful drug is estimated to be 5.5 billion USD (Winegarden, 2014). This number incorporates the cost (incurred by academic and governmental agencies) of failures of the thousands and sometimes millions of compounds that may be screened and assessed early in the R&D process, only a few of which will ultimately receive approval. The overall probability of clinical success (the likelihood that a drug entering clinical testing will eventually be approved) is estimated to be less than 12%.

A worrying trend for pharmaceutical industry is the continued rise in costs of research and development (as seen in figure 2, based on DiMasi, 2014). In addition to regulation, competition in the global market and reduced government funding the size of the field of potential compounds.

Figure Average PhRMA Member Company R&D Expenditures, 1995-2015

The optimal compound to proceed with to the pre-clinical testing phase, given it even actually exists for a specific effect, is the proverbial needle in a haystack (see figure 3) and finding it incurs a significant cost. It is so difficult (and therefore unviable economically) that it is manly funded by academic institutions (universities and public research foundations), governments and philanthropic organizations (DiMasi, 2014).

According to the Tufts Center for the Study of Drug Development (DiMasi, 2014) roughly 30% of the total cost of an approved new compound is attributed to the pre-human phase of the research which includes the drug discovery and the pre-clinical phase. That is a huge potential for savings which can be materialized using the tools and processes described in this paper.

A Nature’s Review article (Waring et al, 2015) presents a model which defines the distinct phases of drug discovery and development starting at the initial stage of “target-to-hit” to the final stage of releasing the drug to the market. The model describes (among other identifiers) the probability of a successful transition from one stage to the next and the phase cost for each project. The model estimates the total cost to achieve one drug launch per year at $1,778 million. per NME launch. It is important to note that this model does not include investments for exploratory discovery research (which as mentioned before is rarely performed in the industry), post-launch expenses or overheads (that is, salaries for employees not engaged in R&D activities but necessary to support the organization).

Figure Candidates Required for a Single Release

1. Convert the Target (a compound involved in the sought pathology that the drug-in-development is intended to act on) to a Hit (a compound which has the desired activity in a compound screen and whose activity has been successfully confirmed).
2. Convert the Hit to a Lead (a confirmed and evaluated compound which has a known dose response, intrinsic activity and other required attributes).
3. Optimize the compound to a sufficiently effective reaction.
4. Pass the preclinical stage at which important feasibility, iterative testing and drug safety data are collected and the validity of the compound is tested to determine the safe dose for human study and assess a product's safety profile.
5. Pass Phase 1 trials (first-in-man studies): the first stage of testing in human subjects mostly aimed at determining the maximum amount of the drug which can be given to a person before adverse effects become intolerable or dangerous.
6. Pass Phase 2 trials which assess how well the drug works.
7. Pass Phase 3 trials which assess the effectiveness of the new treatment (based on the compound) and its value in clinical practice.

Independent researchers such as Paul SM, Mytelka et al (Paul, Mytelka, Dunwiddie, Persinger, Munos, Lindborg, Schacht, 2010) seem to accept the premise and set the focus on the productivity of the process. According to said publication the pharmaceutical industry is suffering from major losses of revenue due to patent expirations, increasingly cost-constrained healthcare systems and ever stricter regulatory requirements. They assert that the key to tackling these challenges such issues pose to both the is to remain within budget limits and substantially increase the quantity and quality of innovative, cost-effective new medicines, contrary to contemporary trends. They present a detailed analysis based on comprehensive, recent, industry-wide data and identify the relative contributions of each of the steps in the drug discovery and development process to overall R&D productivity.

## Machine Learning in Pharmaceutical Research

The secretive nature of pharmaceutical research severely limits the number of publications on the issue. There are a few however.

In 2011 Chapel Hill (University of North Carolina) published “THE QSAROME OF THE RECEPTOROME” (Zhao, 2011) which is a research paper dealing with modeling sets of multiple receptors and its relation with GPCRs. It uses a combinatorial QSAR (Quantitative structure–activity relationship) framework heavily reliant on a Distance Weighted Discrimination (similar to the results of t-SNE) as a score generator and a cost/benefit ratio applied using Decision Trees.

In Application of Predictive QSAR Models to Database Mining Alexander Tropsha discusses a drug discovery strategy that employs variable selection quantitative structure-activity relationship (QSAR) models for chemical database mining.

Same as in the aforementioned publishing the approach starts with the development of rigorously validated QSAR models obtained with the variable selection k nearest neighbor (kNN) method (as opposed to the Decision Trees method mentioned in previous article). Model validation is based on several statistical criteria, including the randomization of the target property (Y-randomization), independent assessment of the training set model's predictive power using external test sets, and the establishment of the model's applicability domain. The most successful models were employed in database mining concurrently and the specific biological activity (characteristic of the training set compounds) of external database entries found to be within a predefined similarity threshold of the training set molecules were predicted based on the validated QSAR models using the applicability domain criteria. Compounds judged to have high predicted activities by all or most of all models are considered as consensus hits.

For the purpose of validating the method the 10 best models were applied to mining chemical databases, and 22 compounds were selected as consensus hits. Nine compounds were synthesized and tested at the NIH Epilepsy Branch, Rockville, MD using the same biological test that was employed to assess the anticonvulsant activity of the training set compounds; of these nine, four were exact database hits and five were derived from the hits by minor chemical modifications. Seven of these nine compounds were confirmed to be active, indicating an exceptionally high hit rate. The approach described in this report can be used as a general rational drug discovery tool.

In article focused on the role of Combinatorial Library Synthesis (Kalliokoski, 2015) the need for dealing with the sheer volume of compounds which can be synthesized. Combinatorial libraries are synthesized by combining building blocks. Many of the commercially available building blocks are too expensive for large scale work and require a reduction of the synthesis space into a more manageable one – potentially by using computational methods.

## GPCRs

The G protein-coupled receptors (GPCRs), as mentioned in the introduction’s application section, have excellent potential for drug discovery with GPCR targets representing up to 30% of the portfolio of many pharmaceutical companies. A GPCR, also called seven-transmembrane receptor or heptahelical (having seven helices) receptor, is a protein located in the living tissue cell membrane that binds extracellular substances and transmits signals from these substances to an intracellular molecule called a G protein (guanine nucleotide-binding protein). GPCRs exist in cell membranes of a wide range of organisms, including plants, microorganisms, and mammals. There are many types of GPCRs with more than 1,000 types encoded by the human genome alone and as a group they respond to a diverse range of substances, including light, hormones, amines, neurotransmitters, and lipids. GPCRs include beta-adrenergic receptors (which bind epinephrine), prostaglandin E2 receptors (which bind inflammatory substances called prostaglandins) and rhodopsin (which contains a photoreactive chemical called retinal that responds to light signals received by rod cells in the eye). The existence of GPCRs was discovered in the 1970s by the American molecular biologist Robert J. Lefkowitz who shared the 2012 Nobel Prize for Chemistry with his colleague Brian K. Kobilka.

A GPCR is made up of a long protein that has three basic regions:

1. An extracellular portion (the N-terminus)
2. An intracellular portion (the C-terminus),
3. A middle segment containing seven transmembrane domains.

Beginning at the N-terminus, the GPCR protein winds up and down through the cell membrane, with the long middle segment crossing the membrane seven times. The seventh part is connected to the C-terminus. When a ligand (a molecule with an affinity for the receptor) is bound by a GPCR, it triggers a conformational change in the seven-transmembrane region of the receptor which in turn activates the C-terminus, recruiting a substance that in turn activates the G protein associated with the GPCR. Activation of the G protein triggers a series of intracellular reactions that ultimately results in the generation of some effect, such as elevated heart rate in response to epinephrine.

Mutations in genes encoding GPCRs can lead to disease in living organisms. As an example, a mutation of rhodopsin could result in congenital night blindness due to continuous activation of intracellular signaling molecules. Worse, acquired mutations in certain GPCRs could lead to cancer by having abnormal increases in receptor activity and expression in cell membranes. The importance of GPCRs in relation to human disease make them useful targets for drug development. For example, antipsychotic activates clozapine and olanzapine block specific GPCRs that normally bind dopamine or serotonin and thus disrupt the neural pathways that lead to pathological schizophrenia. There also exist a variety of agents that stimulate GPCR activity.

# Methodology

In this chapter, we start by describing the needs and requirements the process must meet and analyzing the context in which the process operates. Then we proceed to describe the system model and used statistical parameters.

## Needs

The needs of the pharmaceutical industry for the discovery phase are:

1. The process needs to accept a list of tagged compounds with their respective features.
2. The process needs to accept a significantly larger list of candidate untagged compounds with the exact same features.
3. The process needs to provide a short list of the candidate compounds which are the most likely to have similar results as the tagged compounds.

### Compound Features

The features are common industry characteristics of compounds. For example, a list of features used in one of the experiments included:

5-HT1A, ALogP98, Apol, Formal Charge, Coord Dimension, Is Chiral, LogD, Molecular Weight, Molecular Mass, Molecular Solubility, VSA Total Area, HBA Count, HBD Count, NPlusO Count, Number of Atoms, Number of Bonds, Number of Hydrogens, Number of Explicit Hydrogens, Number of Explicit Atoms, Number of Explicit Bonds, Number of Positive Atoms, Number of Negative Atoms, Number of Spiro Atoms, Number of Bridge Head Atoms, Number of Ring Bonds, Number of Rotatable Bonds, Number of Aromatic Bonds, Number of Bridge Bonds, Number of Rings, Number of Aromatic Rings, Number of Ring Assemblies, Number of Rings3, Number of Rings4, Number of Rings5, Number of Rings6, Number of Rings7, Number of Rings8, Number of Rings9Plus, Number of Chains, Number of Chain Assemblies, Number of Fragments, Number of Complexed Fragments, Number of Metal Atoms, Number of SGroups, Number of Super atoms, Number of Isotopes, Number of Custom Data, Number of Pi Bonds, Number of Repeat Units, Number of V3000Templates, Number of R Group Fragments, Number of Stereo Atoms, Number of Stereo Bonds, Number of Single Bonds, Number of Double Bonds, Number of Triple Bonds, Number of Aliphatic Single Bonds, Number of Aliphatic Double Bonds, Number of Unknown Stereo Atoms, Number of Unknown Stereo Bonds, Number of Dative Bonds, Number of Hydrogen Bonds, Number of Allene Stereo Centers, Number of Atropisomer Centers and Number of Axial Stereo.

The large number of features gives the feature selection process enough space to search in and the number of samples allows the dimensionality reduction to converge on the axis of the selected dimensions.

### IC50

The measure which is used for determining the class of a compound is called half maximal inhibitory concentration or IC50. It is a measure of the effectiveness of a compound in inhibiting a specific biological or biochemical function and quantifies the amount of a particular drug or other substance (inhibitor) which is needed to inhibit a given biological process by half.

IC50 is an often used measure in pharmacological research and therefore is suitable for our purposes.

### Database Creation

The compound database is the result of merging 5 databases of GPCR families together. Those databases contained compounds tagged for their effects on Dopamine (1705 compounds), Histamine (1714 compounds), Serotonin (2762 compounds), Adrenoceptor (637 compounds) and Muscarinic (458 compounds) receptors.

The databases were merged exclusively – meaning that any compound which appeared in more than a single database was discarded. All features were shared were shared by the databases so no features had to be dropped from the resulting database. The result database contains 7276 compounds with 135 features.

### Tagging

The tagged compounds carry the information of the effect of the compound. The compounds in the database are tagged with the organic compound they effect which are:

#### Dopamine

Dopamine is an organic chemical that is excreted by individual cells and plays several important roles in both the brain and the body. In the brain, dopamine functions as a neurotransmitter which is a chemical released by nerve cells to send signals to other nerve cells. One of the few distinct dopamine pathways in the brain plays a major role in reward-motivated behavior and most types of rewards (winning a bet or sugar rush) increase the level of dopamine in the brain. Many addictive drugs increase dopamine neuronal activity.

Dopamine functions outside the central nervous system primarily as a local chemical messenger. It inhibits norepinephrine release in blood vessels; it increases sodium excretion and urine output in the kidneys and reduces insulin production in the pancreas. Dopamine reduces gastrointestinal motility in the digestive system, and reduces the activity of lymphocytes in the immune system

The Dopamine system is associated with several important diseases of the nervous system and some of the key medicinal compounds used to treat them work by altering the effects of dopamine. Parkinson's disease, schizophrenia and attention deficit hyperactivity disorder (ADHD) are associated with unregulated dopamine activity.

#### Adrenoceptors

Adrenergic receptors (or adrenoceptors) are a class of cell receptors that are targets of the catecholamines, especially norepinephrine (noradrenaline) and epinephrine (adrenaline). An catecholamine binding to the receptor will generally stimulate the sympathetic nervous system which causes includes dilating the pupils, increasing heart rate, mobilizing energy, and diverting blood flow from non-essential organs to skeletal muscle (a fight or flight response).

#### Histamine

Histamine is an organic compound taking part in local immune responses as well as regulating physiological function in the gastro intestines and acting as a neurotransmitter for the uterus and is involved in the inflammatory response. Histamine increases the permeability of the capillaries to white blood cells and some proteins, to allow them to engage pathogens in the infected tissues and is produced by basophils and by mast cells found in nearby connective tissues.

#### Muscarinic

Muscarinic acetylcholine receptors are acetylcholine receptors in the cell membranes of certain neurons and other cells. Muscarinic acetylcholine receptors act as the main end-receptor stimulated by acetylcholine released from postganglionic fibers in the parasympathetic nervous system.

Muscarinic receptors are more sensitive to muscarine than to nicotine and many drugs and other substances (for example pilocarpine and scopolamine) manipulate these two distinct receptors by acting as selective agonists or antagonists.

#### Serotonin

Serotonin is a monoamine neurotransmitter that can be biochemically derived from tryptophan. Serotonin is primarily found in the gastrointestinal tract (GI tract), blood platelets, and the central nervous system (CNS) of animals, including humans. It is generally accepted as a contributor to feelings of well-being and happiness.

Serotonin is used to regulate intestinal movements as well as regulate moods, the appetite and sleep. Serotonin affects cognitive functions, including memory and learning. Modulation of serotonin at synapses is thought to be a major action of several classes of pharmacological antidepress ants.

The system is required to apply the tags mentioned above with as high a probability as possible on the untagged compounds.

## Operational Requirements

The system (limited to the scope of the current work), however complex in its implementation, has a very simple operational requirement to fulfill which is:

***The system shall analyze large corpus of compound data and generate the optimal visualization and feature set with regard to the applicability of analyzed compounds as candidates for preclinical trials.***

## Context Analysis

### Corpus

The system operates within the context of a pharmaceutical company with a pre-existing corpus of analyzed compounds and a pool of potential compounds from which to select candidates for preclinical trials. The corpus of the compounds must be in a size (number of compounds \* number of features) which will both provide the algorithm with enough data to provide meaningful results and small enough for the algorithm to converge on a solution in a frame of time which allow the researchers to use it as a decision support system.

### Timeliness

The system is to be used as a non-real-time support system. This means it will perform its function in parallel to the function of an active research and development team and augment its capabilities with decision support capabilities.

### Personnel and Infrastructure Qualifications

The system is to be used by specifically trained personnel so it has no strict user experience limitations on the learning curve it incurs.

The system, being software based, is agnostic to the hardware it runs on and can be executed in various environments such as cloud environment or on-premise environment.

## System Model

The bottom level capabilities of the system (described graphically in Figure 2) are as follows:

### Dimensionality Reduction

In the context of our system the dimensionality reduction serves dual purpose. In addition to making the data more comprehensible it allows for the distance between compounds to be processed in a way which puts emphasis on closeness, which I critical to the system which uses a nearest neighbor as a selection metric.

### Quality Factor

The ability to attribute a certain mapping with a quality factor is critical to determine whether the results can be counted on. There are two processes which need to be measured for quality for the process to be successful:

1. Clustering: this is measured by the relative change in the distances between points – points ***a*** and ***b*** which were closer together than points ***c*** and ***d*** in the original space should be closer in the mapped lower dimension space.
2. Classification: This is measured by the probability of a tagged point having a nearest neighbor with the same tag.

### Progressive Filtering

The projection created for certain sets of features should be progressively filtered so better and better feature sets are used.

Figure Capabilities

The system needs to be able to measure the distance between compounds effectively and for that purpose needs to intelligently reduce the dimensions of the data space (as opposed to simply assuming equal weight on each dimension) and use the L2 distance. In addition, the system needs to measure the quality of the dimensionality reduction by using clustering algorithm on a-priori tagged data.

The system needs to select the tested features of the data out of a large field of diverse features. For that purpose, the feature selection algorithm works in conjunction with an optimization engine using genetic algorithms.

The designed system is currently demonstrated using a combination of MATLAB (preforming pre-processing and the enveloping optimization process) and C++ code performing the dimensionality reduction process.

As mentioned above the system’s goal is identify the optimal set of features by which the visual separation between active and inactive compounds will be maximal (by parameters described below). Using the optimal set of features, we could in the future analyze unknown compounds and determine with a high probability their activation status.

The purpose behind the tool is to identify potentially effective compounds using previously known information about a small number of compounds. For example, if we can tag a few of the compounds we can – by association – estimate with a high probability of success whether the compounds near them in the resulting map (such as in Figure 6 Compounds grouped by bitterness) have high effectiveness potential.

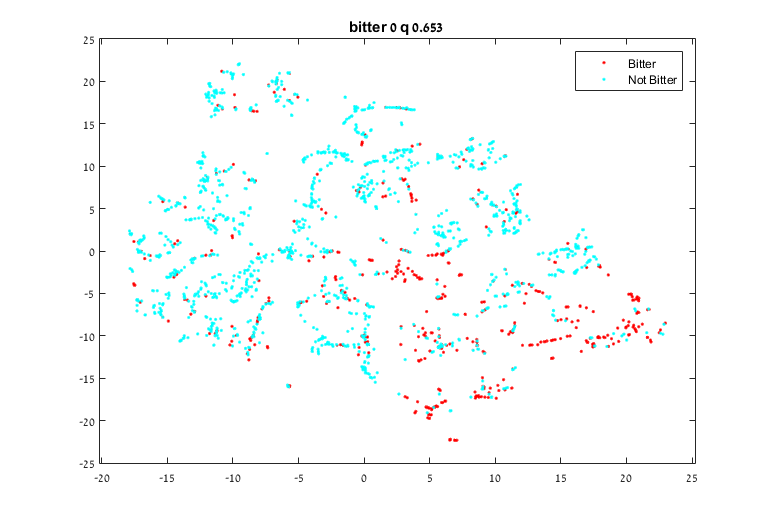


Figure Compounds grouped by bitterness

Following is a graphical depiction (Figure 6 t-SNE Optimization Algorithm) of the system meeting the requirements described above. The system is comprised of 5 functional parts: input database, feature selection, dimensionality reduction, fitness function calculation and Model selection and generation using genetic algorithms to create ever improving generations until a quality condition is satisfied or the maximal number of iterations is reached.

Input Database

Selection of Features

Run t-SNE

Optimization Engine (GA)

Calculate Fitness Function

Select Best Model

Figure t-SNE Optimization Algorithm

### Input Data

The first stage consists of data input consisting of GPCR families as described in the literature review. The system accepts digital comma separated value files created using specifically prepared data set (compiled from private data sources) and, for some, classifications into their compound tagging group.

### Feature Selection

Under the assumption that some of the features carry more information than others for the snapshot of the world the data represents the goal of the algorithm is to select the optimal feature set. The selected feature selection optimization algorithm was Genetic Algorithm.

#### Genetic Algorithm

In optimization theory, ***genetic algorithm*** is a natural selection based method for solving both constrained and unconstrained optimization problems. The genetic algorithm iteratively modifies a population of possible individual solutions. At each iteration, the genetic algorithm selects individuals from the current population according to certain selection, mutation and crossover rules (addressed below) and uses them as parents to produce the children of the next generation. Each and every successive generation brings the population closer to an optimal solution. Genetic algorithms can be applied to a variety of optimization problems that are not well suited for standard optimization algorithms, including problems in which the objective function is discontinuous, nondifferentiable, stochastic, or highly nonlinear.

A genetic algorithm uses three main types of generation rules at each step to propagate the next generation from a current population:

1. **Selection** rules select the individuals, called parents, that contribute to the population at the next generation.
2. **Crossover** rules combine two parents to form children for the next generation.
3. **Mutation** rules apply random changes to individual parents to form children.

In the context of the compound selection system, as the basis for a genetic algorithm the initial population is 20 sets of randomly selected features. From each generation, the next generation is created as follows:

1. The fittest 30% are carried as they are to the next generation
2. Mutations of the fittest 20% are generated into the next generation
3. Crosses of the fittest 50% are generated into the next generation.

After the process stops improving (given at least 3 iterations without improvement) it’s assumed that the optimal feature set has been found. Otherwise the process will stop at the maximal number of iterations (which is set to 128).

### Dimensionality Reduction

We chose to use t-distributed stochastic neighbor embedding (t-SNE) for dimensionality reduction. Unlike more commonly used dimensionality reduction algorithms (such as PCA) which are mainly concerned with preserving large pairwise distances t-SNE maintains structure by using metrics of nearest neighbor as opposed to distance.

#### t-SNE

***t-distributed stochastic neighbor embedding*** (t-SNE) is a nonlinear dimensionality reduction algorithm that is especially designed for embedding high-dimensional data into a space of two or three dimensions, which can then be visualized in a scatter plot. For the purpose of clustering it models each high-dimensional object by a two- or three-dimensional point in such a way that similar objects are modeled by nearby points and dissimilar objects are modeled by distant points.

The t-SNE algorithm has two main stages:

1. t-SNE constructs a probability distribution over pairs of high-dimensional objects in a way that makes similar objects have a high probability of being selected and dissimilar points have an extremely small probability of being selected.
2. t-SNE defines a similar probability distribution over the points in the low-dimensional map, and it minimizes the Kullback–Leibler divergence (a measure of how one probability distribution diverges from a second expected probability distribution) between the two distributions with respect to the locations of the points in the map.

In the domain of compound tagging, it is critical that the process of preserving large distances (which carry little data) will not affect preserving local (short) distances since the shortest distance estimation along the selected features is what determines the nearest neighbor and for our purpose of classifying untagged samples this is the most important quality.

### Fitness Calculation

The fitness of a feature set (representing the quality of the classification algorithm) has been measured using multiple metrics (fully described below) including the nearest neighbor being of the same class, two out of three nearest neighbors and three out of five nearest neighbors. All metrics behave in a similar manner so finally the fitness score is determined by the percentage of tagged samples having a nearest tagged neighbor with the same tag.

Because there are datasets where the target tag is a very significant minority simply applying nearest neighbor counting as a measure of quality can create a false high quality measures. For this reason only the positively tagged (the tag which we are interested in finding) are taken into consideration and not the general population.

The trust metric (representing the quality of the clustering algorithm) indicates that the untagged neighbors are also neighbors in the higher dimensional space. This indicates how much the structure of the data has persisted through the dimensional reduction.

### Model Selection

The model with the best fitness score is selected for a dataset. For a specific tag the untagged compounds with the highest percentage of neighbors that share that tag are identified as the best candidate compounds for preliminary pre-clinical tests



## Statistical Parameters for Evaluation

Three statistical parameters test the success of the and a control measure is used for verification purposes:

**Single neighbor quality:**  this is the main quality parameter and is calculated in the following manner: For each sample of the positively tagged compounds the value of the single neighbor quality measure is true if the nearest neighbor is also positively tagged and false otherwise. This statistic answers directly the following question: If an untagged compound is the nearest neighbor of a positively tagged compound – what is the probability that it is also carries the characteristic of positively tagged compounds?

**Three neighbors quality:** this is the second quality parameter which answers a similar question: If an untagged compound has two of its three nearest neighbors positively tagged compounds – what is the probability that it is also carries the characteristic of positively tagged compounds?

**Five neighbors quality:** this is the tertiary quality parameter which answers a similar question: If an untagged compound has three of its five nearest neighbors positively tagged compounds – what is the probability that it is also carries the characteristic of positively tagged compounds?

**Trust level:** this is a control measure describing the preservation of high dimensional structure at two dimensional space. This parameter is used for control because for higher levels of k-neighbor quality we expect the trust to be lower – since the structure is disassembled to reveal the differentiating parameters.

# Results

## Testing

### Test Methodology

In order to test the system smaller real-world datasets, from the pharmaceutical domain as well as from the financial domain, were used. These data sets are simple enough and small enough to be able to visually determine the success of the system in separating the dataset in accordance with the compounds’ tagging.

Since the data is separate from the tags and the model is not persistent there is no need to perform cross validation and we can use the entire span of the data to generate the results.

### Result Information Parameters

In all of the following example the images are presented with their result information:

Table Result Parameters

|  |  |
| --- | --- |
| **Name:** | The name of the dataset. |
| **Separated feature:** | The name of the sought-after tag. |
| **Single neighbor quality:** | The quality calculated by a single nearest neighbor. |
| **Three neighbors quality:** | The quality calculated by two out of three nearest neighbors. |
| **Five neighbors quality:** | The quality calculated by three out of five nearest neighbors. |
| **Trust level:** | The clustering quality metric. |
| **Selected features** | The selected features encoded in a hexadecimal string. |

The features are encoded binarily and displayed in hexadecimal form with each letter representing four features. For example if we have the following ten features 0110101011 they will be represented as 1AB.

### Random Dataset

For the sake of completeness, a random dataset was generated (500 samples of 24 features tagged 0 and 1) and analyzed by the system. Since the system employs intelligent feature selection it was expected that the resulting map, utilized by the features best fitting the tags, would result in a slightly better than random quality score.

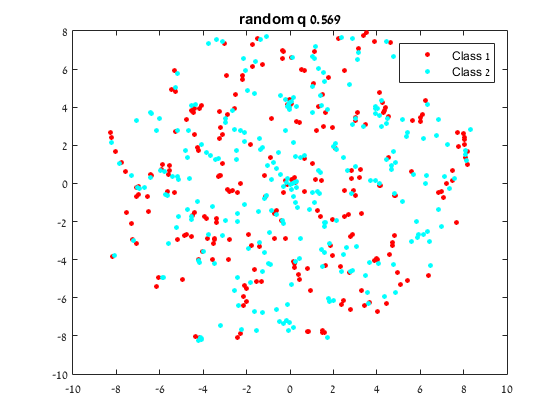


Figure random sample test

As seen in in the resulting projection map (Figure 3) the quality is 0.569 – meaning that for any sample the probability of having the neighbor with the same tag is 0.569, which is as expected.

Table Random Dataset Results

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | *Q-1* | *Q-3* | *Q-5* | *Trust* |
| *Random* | 0.569 | 0.498 | 0.519 | 0.636 |

### Bitterness Dataset

The bitterness database consists of 2074 compounds with 19 features and their bitterness quality. We can see the result (Figure 4) that there doesn’t appear to be a very strong correlation between the features and the bitterness level even at the optimal feature selection and only 0.64 of the bitter compounds were mapped near a nearest neighbor that is also tagged as bitter.

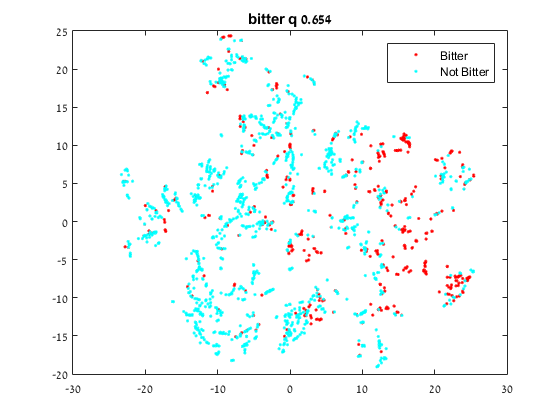


Figure Bitterness Results

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | *Q-1* | *Q-3* | *Q-5* | *Trust* |
| *Bitter* | 0.653 | 0.616 | 0.578 | 0.128 |

## GPCR

The GPCR database contains the 7276 samples detailed above. Table 2 Result Summary presents the results obtained by the t-SNE optimization algorithm for the five GPCR families. For each family a model was performed to locate the feature space that would provide the best separation between the family and the rest. Overall, the t-SNE algorithm led to models with good statistical parameters. For each family between 93.8% and 99.3% of the total samples had a nearest neighbor in the 2D representation from their family. In addition, between 93.4% and 99.3% of the total samples had two nearest neighbors out of three in the 2D representation from their family, and finally at each family between 92.1% and 99.3% of the total samples had three nearest neighbors out of five in the 2D representation from their family.

### Result Summary Table

The following table presents the best results for each target and the relevant quality parameters of quality and trust. The bottom line is the average quality parameters for ll the targets.

Table Result Summary

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | *Q-1* | *Q-3* | *Q-5* | *Trust* |
| *Dopamine* | 0.9650 | 0.9650 | 0.9610 | 0.0870 |
| *Adrenoceptors* | 0.9830 | 0.9850 | 0.9840 | 0.0770 |
| *Histamine* | 0.9670 | 0.9630 | 0.9640 | 0.0780 |
| *Muscarinic* | 0.9930 | 0.9930 | 0.9930 | 0.0760 |
| *Serotonin* | 0.9380 | 0.9340 | 0.9210 | 0.0900 |
| *Average* | *0.9692* | *0.9680* | *0.9646* | *0.0816* |

### Results Analysis

#### Dopamine

Compounds associated with Dopamine are 1705 out of 7276 compounds.

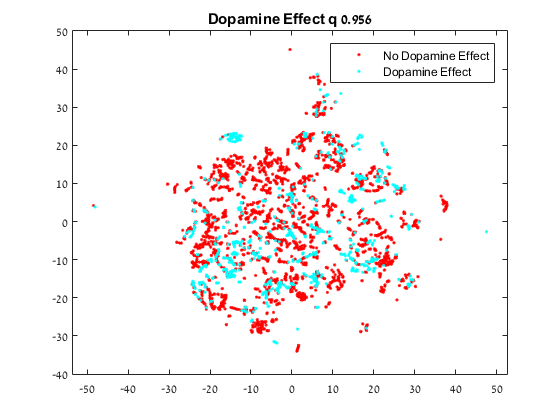


Figure Dopamine Results

The low trust level demonstrates the high level of the algorithm aggressiveness – compounds which were close in the high dimensional space were not close in the resulting mapping.

The high-quality level though indicates that the system was successful in clustering the similarly tagged compounds together.

As seen in Figure 6 Dopamine Zoom In) We can see that seemingly single dots (representing compounds without near positive tagged compounds) are small tight clusters.

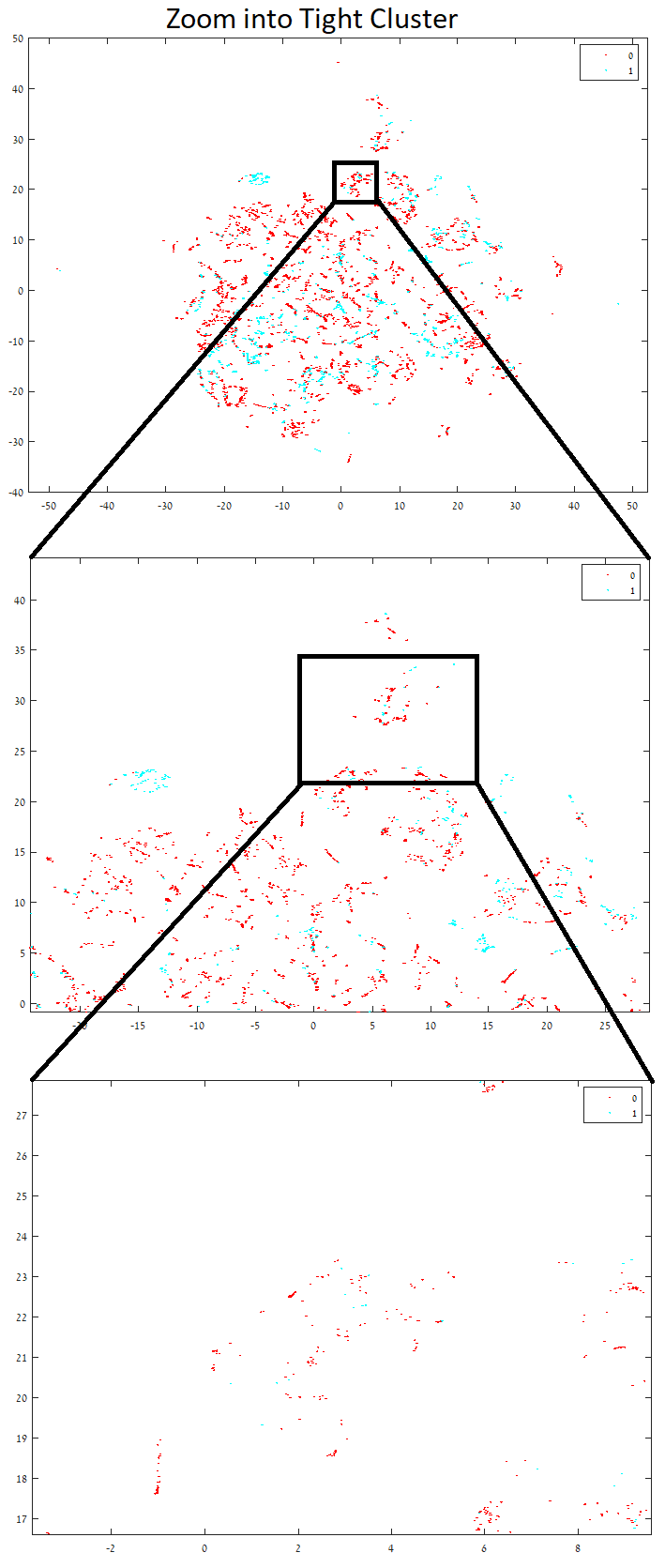


Figure Dopamine Zoom In

#### Adrenoceptors

Compounds associated with Adrenoceptors are 637 out of 7276 compounds.

Much like the Dopamine results the low trust level demonstrates the high level of the algorithm aggressiveness and also like the Dopamine the even higher quality level though indicates that the system was successful in clustering the similarly tagged compounds together (Figure 7).

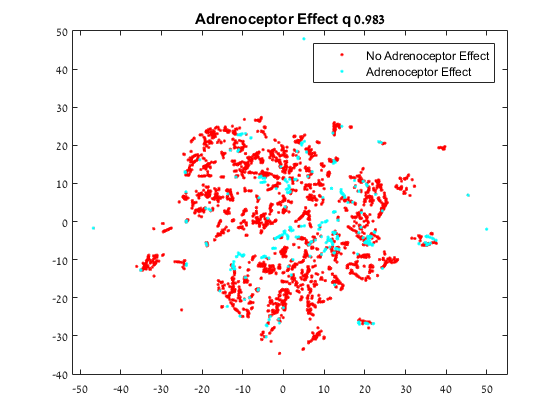


Figure Adrenoceptor Results

#### Histamine

Compounds associated with Histamine are 1714 out of 7276 compounds.

Tagging Histamine compounds (Figure 8) shows similar results to tagging Dopamine.

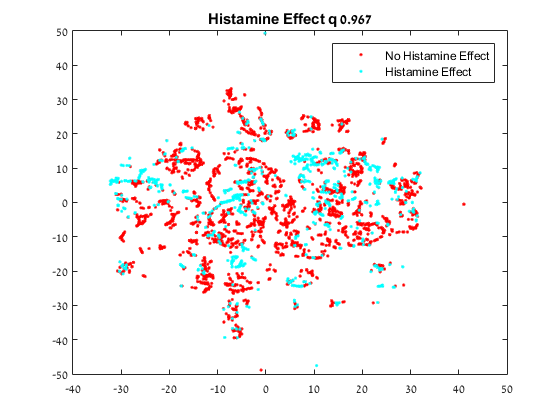


Figure Histamine Results

#### Muscarinic

Compounds associated with Muscarinic are 458 out of 7276 compounds.

Tagging Muscarinic compounds (Figure 9) shows excellent results. Similar to the test performed on qualitative financial ratios the scope of the map the visual indication seems to contradict the quality figures but it bears reiterating that the image represents more than 7000 compounds. Taking the single compound in the middle (for example) and zooming into the map we can see (Figure 10) that it is in fact a cluster of two compounds bunched together in a manner which makes it seem like a single compound in a compressed space.

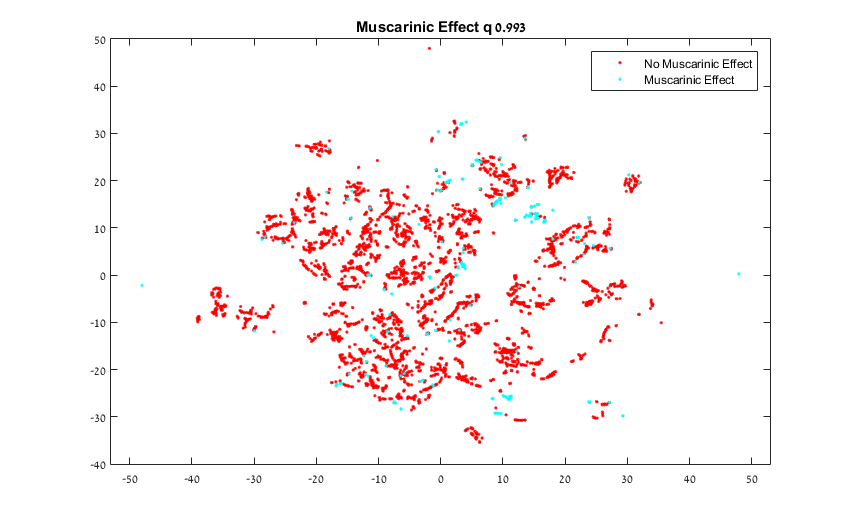


Figure Muscarinic Results

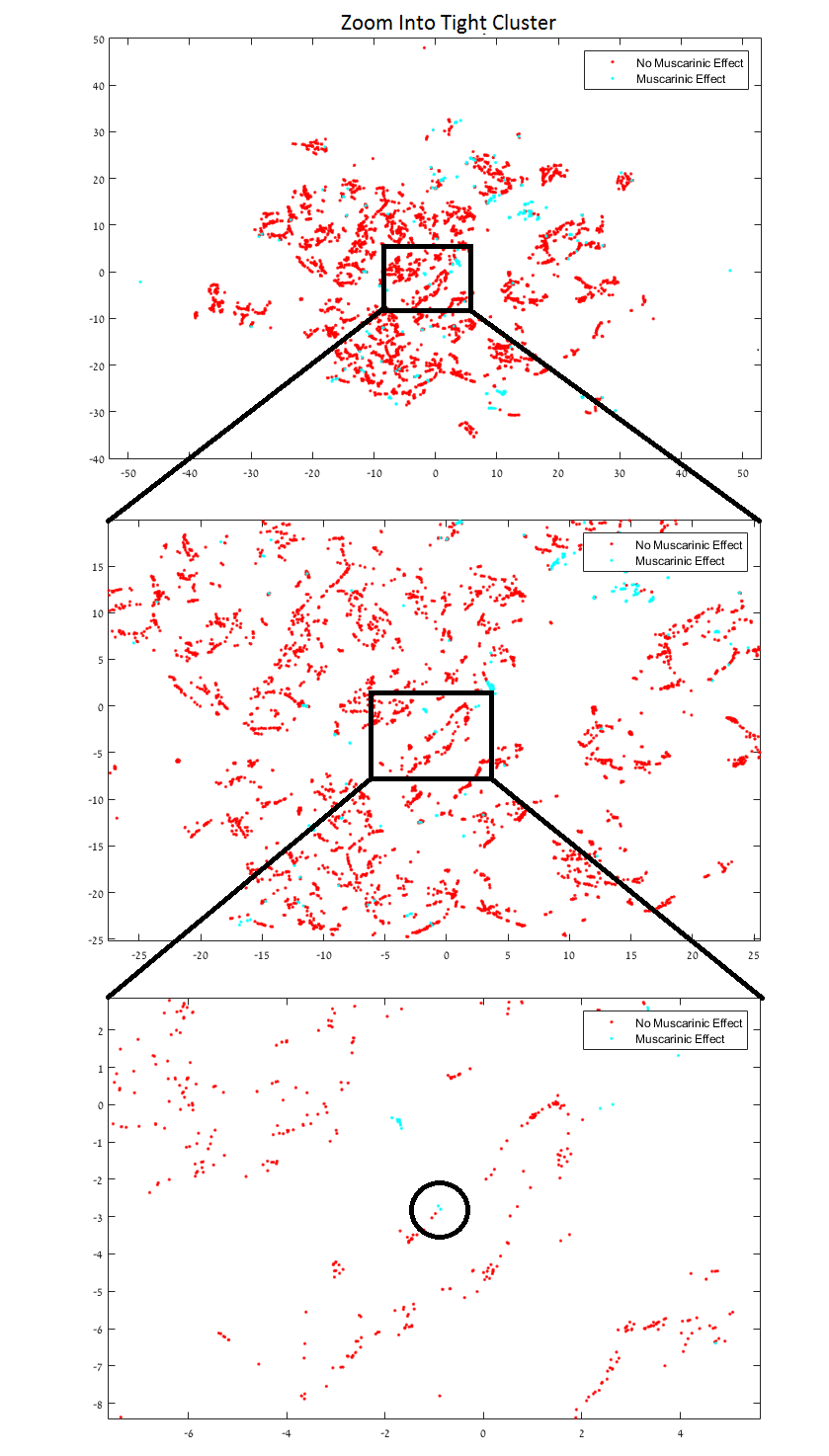


Figure Muscarinic Results Zoom In

#### Serotonin

Compounds associated with Serotonin are 2762 out of 7276 compounds.

As the most dominant effect in the dataset (with regard to compound count) Serotonin has the lowest quality score (Figure 11) which is still better in orders of magnitude than random selection.

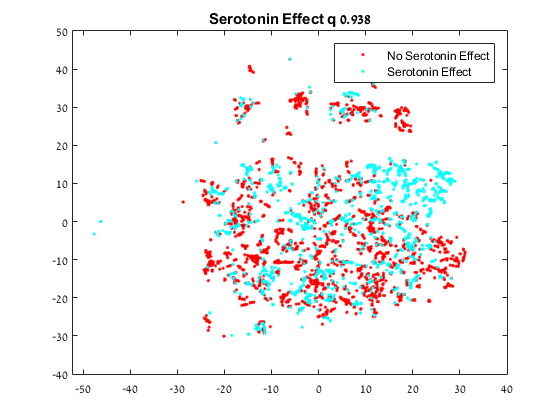


Figure Serotonin Results

# Discussion

The algorithm was designed to provide us with a classification of the compounds with respect to an effect on a specific biological activity (IC50). The value of the algorithm is measured by its success in correctly partitioning the database of compounds into active and inactive compounds with concerning each biological activity.

The measures of partition used to evaluate the algorithm are (as mentioned above) the portions of the positively tagged compounds (meaning affecting the specific agent) with K out of N neighbors having the same tagging, K and N being (1,1), (2,3) and (3,5).

On average the results (97.7% accuracy for 1NN, 97.65% accuracy for 3NN and 97.55% for 5NN) indicate that we have a very good classification process. The average trust level of 7.95% indicates that the process selected the features in a manner which was not linearly related to the original multi-dimensional representation of the data.

We have demonstrated that it is possible to classify families of GPCRs For example, if we are interested in finding new Histamine inhibiting molecules in a database of unanalyzed molecules we follow these steps:

1. Add known Histamine inhibiting molecules to the database.
2. Execute the process.
3. Pick the molecules closest to the known Histamine inhibiting molecules as the most likely to be Histamine inhibiting molecules as well.

Moreover, the process provides us with the selected features so as long as the context of the process remains static we only need to employ the dimensionality reduction algorithm for compound selection.

# Conclusions

## Conclusions

We have demonstrated the research hypothesis discussed in this paper and used computational visualization method to classify an industry class database into the desired classes.

Examining the result for all the tested effect show that in all the cases the system produced results that were superior to random selection by at least one order of magnitude. It remains to be seen whether that would be sufficient for the industry to start including the discovery phase as an integral part of their R&D process.

Compared to the benchmark introduced above we have exceeded our expectations and we can see that in most cases no more than seven percent of the initial given compounds were left for the costly process of physical compound testing – by which we reduced the potential cost of the discovery phase to less than ten percent of its original cost.

Using the process described above we can (for example) discard any of the untagged compounds which nearest neighbor is either tagged negatively or untagged. In the GPCR database that would discard the vast majority of the compounds and leave us only with the prime candidates for testing.

## Future Work and Research Limitations

There are a few improvements that can be made to the system to make it more suitable to work in the industry.

While the execution time is still a concern the system upon which the research was conducted on is a single home personal computer. The most trivial means of speeding the process would be to execute the dimensionality reduction of each generation member on a different machine and that would speed the process by a factor equal to the size of each generation. More complicated solutions would include multi core implementations of t-SNE.

These solutions, even though potentially speeding up the process by a factor of at least 160, might prove insufficient since the t-SNE algorithm has both time and space quadratic complexities. Even with industrial power machines and cloud computing databases of 100000 entries might have unrealistic computing power requirements – which means a powerful pre-processing will be required or a variant of t-SNE with lower complexity (there are some ongoing research efforts into that objective) will be used.

### Dataset Size

The dataset which the experiment was conducted with is a real industry standard dataset with regards to features. It is expected however that the pharmaceutical companies have datasets in one or more orders of magnitude larger. Since both the computational and the memory complexity of t-SNE are O(n4) larger datasets will not be manageable on a single machine and (as mentioned above) a distributed algorithm will have to be utilized.

### Fast Dimensionality Reduction

There are a few ongoing research efforts into creating a faster version of t-SNE which would be able to handle significantly larger databases. Additionally there are other dimensionality reduction algorithms which might prove a reasonable substitution.

# Commercial System Plan

This chapter includes a detailed discussion of applying the algorithm described in this paper within the context of a pharmaceutical firm with the data and resources to take advantage of the potential results.

## Assumptions

To plan the system, we would have to make a few reasonable assumptions. Following are the assumptions and their rationale.

### Budget

As described above, the currently estimates cost of the discovery phase, which results in finding enough viable candidates to bring a single compound to market, is 1.25 billion USD and is too high for the pharmaceutical companies.

Without access to full marketing and financial data it is difficult to accurately evaluate how much financial resources would be a reasonable investment for a pharmaceutical company. It seems reasonable to pick a number one order of magnitude smaller than current estimated cost so the number of 100 million USD per drug will be used. Furthermore, additional XXXX will make a reasonable setup cost.

### Database Size

As was mentioned above it takes a corpus of 125,000 candidates to produce 75 viable pre-clinical trials candidates out of which a single drug is released to the market (Winegarden, 2014).

It is therefore assumed that a commercial database will be of size 125,000 – one order of magnitude larger than the one used to obtain the results in this paper – with a similar number of features per compound.

### Algorithm

The algorithm as presented in this paper has a complexity of . On the experiment database on a single powerful PC analysis takes roughly an hour. A database 16 times larger would take roughly times to execute which amounts to about 7 and a half years of computing time. There are 2 steps required to bring that number down to a practical number:

#### Quicker Algorithm

There are flavors of t-SNE which have similar results but have a complexity of only . Using such an algorithm will reduce execution time in our case by of which would take the execution time down to about 2600 hours.

#### Parallelization

Most of the time in the algorithm is spent on performing dimensionality reduction on single instances of selected features. Since these computational tasks are completely independent they can be performed in parallel, reducing the running time of the algorithm to hours.

#### Powerful Hardware

Even though 90 hours is probably an acceptable execution time for the algorithm it is possible to use especially strong processors. The tests for this paper were performed on Intel(R) Core(TM) i7-6700HQ CPU @ 2.60GHz. Advanced CPUs such as Intel Xeon E5 2697 v3 operate at little over 3 times higher Million Instructions per Second (MIPS) score and would allow the algorithm to complete a full cycle in roughly 24 hours. For the same of comparing options the assumption would be that the requirement is that the process will repeat daily so the stronger CPUs are a must.

## Needs Analysis

To plan the

The commercial needs of the pharmaceutical industry were discussed above in the needs (‎3.1) section. The operational requirements are as follows:

### Operational Requirements

1. The system shall analyze databases of at least 100,000 partially tagged compounds with at least 130 features per compound.
2. The system shall tag as candidates the untagged compounds that are most likely to share the desire properties as the tagged compounds.
3. The system shall provide a human readable list of candidate compounds ordered by their estimated likelihood to share the desired properties with the tagged compounds.
4. The system shall generate a list from a database within 24 hours.
5. The cost of each list generation shall be up to 3,000 USD.
6. The setup cost of the system shall be up to 5 million USD.

### Concept Exploration

The top-level process (shown in Figure 12) shows the main participants in the entire flow:

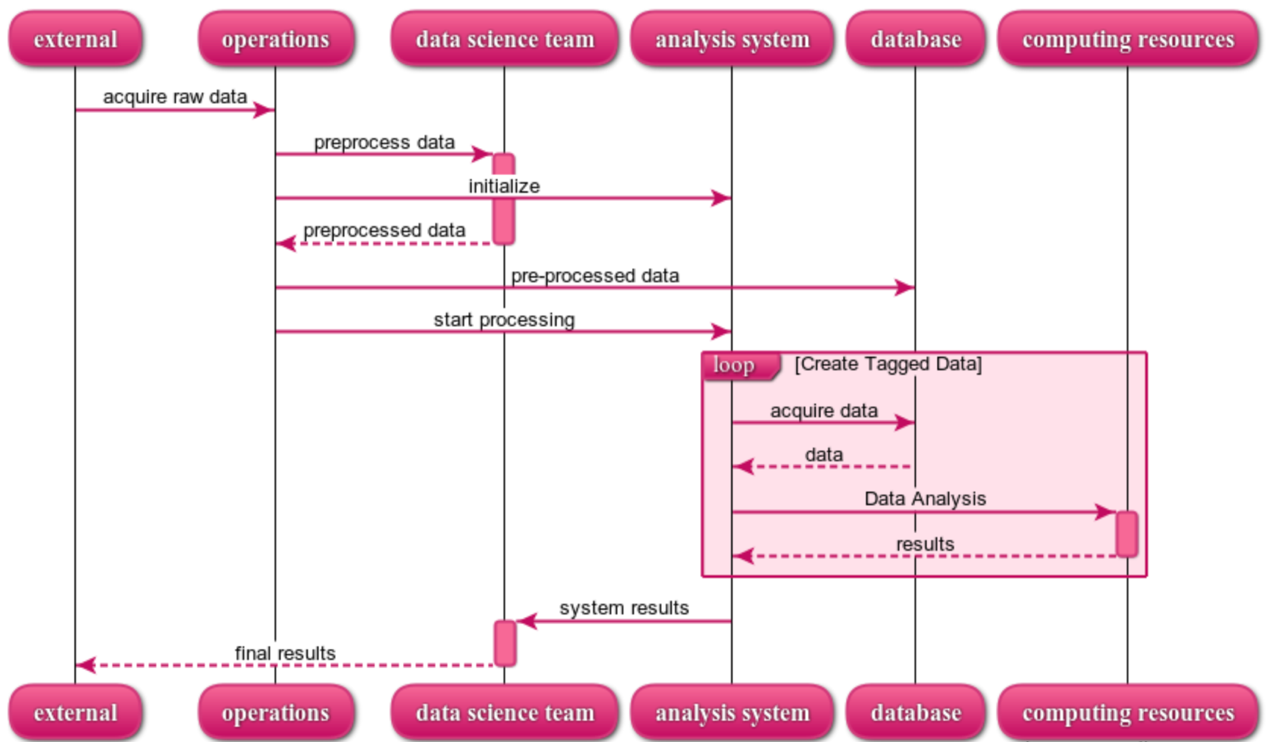


Figure Top Level Process

The participants are:

#### Operations

The management and corporate team which facilitates the operations of the system within the corporate structure.

#### Data Science Team

A team of professional data scientists responsible to provide data pre-processing and final results analysis capabilities.

#### Analysis system

An automated system managing the entire flow of the tagging process by consuming computing resources to analyze the data in the database.

#### Database

A persistence mechanism for the data corpus.

#### Computing Resources

The resources which provide the storage and computing power required for the analysis process.

There are 2 concepts to explore for the system: on premises and cloud based. Following is a description of each and an analysis with respect to cost (amortized over two years), scalability and recoverability. As the impact of the database specification is negligible compared to the computing power the algorithm requires the option analysis will focus on the CPU and memory requirements.

#### On Premises

The computing resources could be deployed internally. A set of 30 CPUs will be utilized for that purpose and their life expectancy – due to the constant usage – would be 2 years. As one CPU costs about 2800 USD to acquire total purchase, including peripheral and hosting equipment would be roughly USD. The amortized cost of cooled and monitored storage room and the IT personnel required to maintain the hardware would add 5,000 USD per month to an amortized total of:

USD

With 24 being the number of periods (months) and 0.012 being the monthly internal rate of return. The total amount in present value is little under 200,000 USD.

Scaling an on premises deployment is not trivial. Scaling up means purchasing more machines and allocate more space which might not be available in a short notice. Scaling down would mean putting machines back in storage (since selling back into the market is not economic) and potentially relocating to a smaller space.

Recovering the cost in case of project termination is not trivial either. As mentioned above selling machines back to the market will mean recovering up to 75% of their original cost so it might make sense to store them for future project which might require powerful computing capabilities.

#### Cloud Hosting

Consuming the same amount of computing power on Amazon requires using the c4.8xlarge instances which cost 1.591 USD per hour or roughly 1,150 USD per month using on-demand pricing. Using the same figure for previous option we get:

USD

Using spot pricing (which means using free instances at discount rating) at an estimated discount rate of 70% brings the price to just USD.

Scaling a cloud deployment, it very easy and simply requires the termination of the machine subscription on the cloud platform. Since the cost of the machines is paid monthly there is a little to no issue of recovering costs since the maximal period wasted would be one month which is roughly 4% of the cost.

### Selected Concept

Both concepts are compared using AHP.

#### Criteria Weights

The prioritization of criteria is displayed in figure 13. Since the project is priced at the very low end of pharma projects the criteria of controlling costs and the ability to scale up or down are deemed more important.

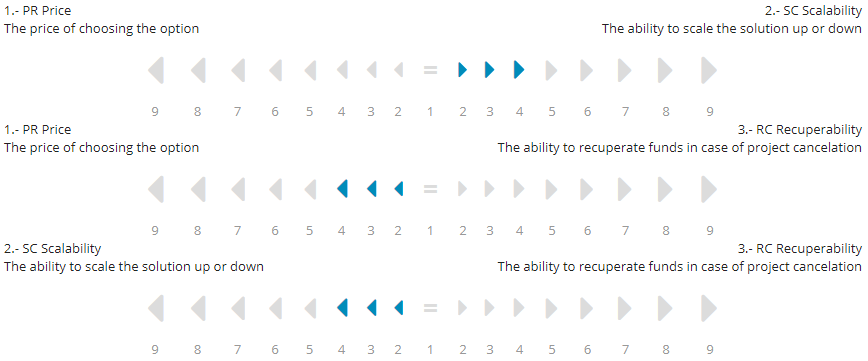


Figure Criteria Prioritization

#### Final Result

The final results are displayed in Table 4 Deployment Options Comparison.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | Price | Scalability | Recoverability | **Result** |
| Weight | 0.27 | 0.62 | 0.11 |  |
| **On Premises** | 0.8 | 0.13 | 0.13 | **0.31** |
| **Cloud Computing** | 0.2 | 0.87 | 0.87 | **0.69** |

Table Deployment Options Comparison

The optimal concept to implement is the cloud computing concept.

## Risk Management

Being a research-intensive project the algorithm’s commercial implementation is risky by nature. The inherent risks should be identified, quantified and tackled according to their impact on the project. The known risks are:

### Fast Implementation (RS1)

The entire premise of the implementation is based on the assumption that the algorithm with complexity of will be implemented and be commercially available. It is a reasonable assumption given the nature of the research involved but a failure will be catastrophic since the algorithm currently cannot analyze commercial sized databases in a reasonable time.

### Data Availability (RS2)

It is possible that there will not be enough readily available tagged data without an additional tagging effort and the tagging effort might prove too costly. This will slow the system’s operations and might even grind it to a halt.

### System Usability (RS3)

The benefit of the system is heavily reliant on the ability of a data science team to interpret the results and successfully hand it over to the R&D teams who need to validate the discovered compounds and approve those compounds for pre-clinical trials.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Likelihood | 5 | 0 | 0 | 0 | 0 | 0 |
| 4 | 0 | 0 | 0 | 0 | 0 |
| 3 | 0 | 0 | 1 | 0 | 1 |
| 2 | 0 | 0 | 0 | 0 | 0 |
| 1 | 0 | 0 | 1 | 0 | 0 |
|  | 1 | 2 | 3 | 4 | 5 |
|  | Consequences | | | | |

Table Risk Map

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **ID** | **Rating** | **Name** | **Likelihood** | **Consequence** | **Status** |
| RS1 | High | Fast Implementation | Medium | Severe | Open |
| RS2 | Low | Data Availability | Low | Moderate | Open |
| RS3 | Medium | System Usability | Medium | Moderate | Open |

Table Risk Impact

### Risk Management Plan

The analysis suggests that before the initial efforts to create the system commence the only severe risk. the algorithm for faster t-SNE, need to be fully resolved. The other risks can be addressed while the system is being deployed. The risk management plan will be described in the deployment plan section.

## Setup and Deployment Plan

Drawing on the results of previous sections the setup and deployment plan would be as follows in the following Gantt chart (Figure 14). Each phase depends on the success of the previous phase and cannot start without the previous phase concluding successfully.

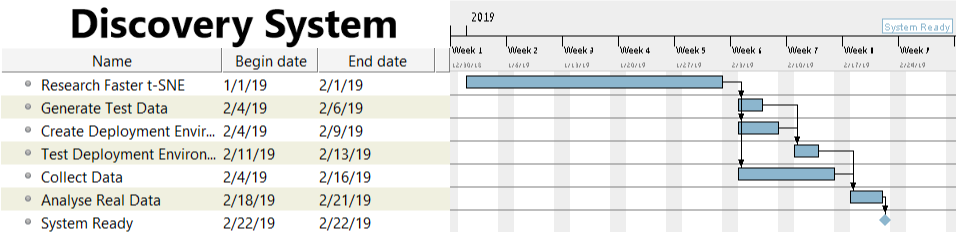


Figure Setup Gantt Chart

## Testing and Evaluation

This paper demonstrates – in a smaller scale – how the system should be tested. Tagged data with known qualities should produce clear cut classification while randomly generated data should produce poor classification.

## Operations

Following the “System Ready” milestone the system operates in the context of the process described in Figure 12 - Top Level Process. In case of a suggested upgrade to the system a mirroring system can be created easily using the cloud infrastructure the system resides in.

# Conclusions

We have demonstrated the research hypothesis discussed in this paper and used computational visualization method to classify an industry class database into the desired classes.

Examining the result for all the tested effect show that in all the cases the system produced results that were superior to random selection by at least one order of magnitude. It remains to be

# List of Publications

* "The Big Data Challenge in Drug Design: Data Visualization", Poster presented before the MuTaLig COST ACTION, WG meeting 2016, Budapest (HU), Nov 19-20 2016

# References

1. English, R.; Lebovitz, Y.; Griffin, R., Institute of Medicine (US) Forum on Drug Discovery Development and Translation. Transforming clinical research in the United States. Challenges and opportunities: workshop summary. Washington (DC): National Academies Press (US); PubMed PMID: 21210556 **2010**
2. Joseph A. DiMasi, Tufts Center for the Study of Drug Development, Cost of Developing a New Drug
3. H Geerts; A Spiros; P Roberts and R Carr, Has the Time Come for Predictive Computer Modeling in CNS Drug Discovery and Development? CPT: Pharmacometrics and Systems Pharmacology · November 2012
4. Breakthrough Business Models: Drug Development for Rare and Neglected Diseases and Individualized Therapies: Workshop Summary, National Academies Press
5. An analysis of the attrition of drug candidates from four major pharmaceutical companies: Michael J. Waring, John Arrowsmith, Andrew R. Leach, Paul D. Leeson, Sam Mandrell, Robert M. Owen, Garry Pairaudeau, William D. Pennie, Stephen D. Pickett, Jibo Wang, Owen Wallace & Alex Weir.
6. Wayne Winegarden, Ph.D, The Economics of Pharmaceutical Pricing, Pacific Research Institute.
7. Guiyu Zhao: The QSARome of the Receptorome: Quantitative Structure-Activity Relationship Modeling of Multiple Ligand Sets Acting at Multiple Receptors, University of North Carolina
8. Robas N, O’Reilly M, Katugampola S, Fidock M: Maximizing serendipity: strategies for identifying ligands for orphan G-protein-coupled receptors. Curr Opin Pharmacol 2003, 3:121-126.
9. Flower DR: Modelling G-protein-coupled receptors for drug design. Biochim. Biophys. Acta 1999, 1422:207-234.
10. Paul SM, Mytelka DS, Dunwiddie CT, Persinger CC, Munos BH, Lindborg SR, Schacht AL: How to improve R&D productivity: the pharmaceutical industry’s grand challenge. Nat Rev Drug Discov 2010, 9:203-214.
11. Bockaert J, Pin J: Molecular tinkering of G protein-coupled receptors: an evolutionary success. Embo J. (1999) 18:1723.
12. Drews J: Drug discovery: a historical perspective. Science (2000) 287:1960-1964.
13. Hopkins Al, Groom Cr: The druggable genome. Nat. Rev. Drug Discov. (2002) 1:727-730.
14. Klabunde T, Hessler G: Drug design strategies for targeting G-protein-coupled receptors. Chembiochem. (2002) 3:928-944
15. Tuomo Kalliokoski: Price-Focused Analysis of Commercially Available Building Blocks for Combinatorial Library Synthesis, Lead Discovery Center GmbH (2015).

# Appendix A – Matlab codE

## main.m

The main loop performing the genetic algorithm over the results of the dimensionality reduction using the main quality measure as fitness function.

population\_size = 20;

clusterGen = 128;

local = './large\_files/gpcr.csv';

tags = './large\_files/gpcr\_labels.csv';

name = 'gpcr';

foul\_counter = 0;

%% Read the data source

try

new = csvread(local);

if trial == 3 || trial == 8

data = new(:, 1:end);

fid = fopen(tags,'r');

labels = textscan(fid, '%q');

labels = cellstr( labels{1});

fclose(fid);

else

data = new(:, 1:end-1);

labels = new(:, end);

end

catch ME

continue

end

y = unique(labels);

n = zeros(length(y), 1);

for iy = 1:length(y)

% for cells n(iy) = length(find(strcmp(y{iy}, labels)));

n(iy) = length(find(strcmp(y(iy), labels)));

end

% zero out according to the experiment)

for zeroing = 1 : experimentCount

if experimentCount > zeroing

[~, itemp] = max(n);

n(itemp) = 0;

end

end

[~, itemp] = max(n);

mode = y(itemp);

if trial == 3 || trial == 8

labels = strcmp(labels, mode);

else

labels = labels == mode;

end

[m,n] = size(data);

% labels = labels';e z

%% Process the data

h1 = figure;

t = 6;

p = 10;

q = 0;

old\_q = -inf;

old\_t = 0;

top\_seed = 0;

jumps = 0;

iSeed = 1;

iQuality = 2;

iTrust = 3;

iQuality3 = 4;

iQuality5 = 5;

iMap = 6;

iFeatures = 7;

population = [];

maps = [];

features = [];

populationFeatures = {};

% Create the seed population

for sz = 1 : population\_size % number of samples

[map, q, q3, q5, trust, features, c1, featuremap] = singletsne(new, n, labels);

population{sz, iSeed} = featuremap;

population{sz, iQuality} = q;

population{sz, iQuality3} = q3;

population{sz, iQuality5} = q5;

population{sz, iTrust} = trust;

end

% Sort the existing population

population = sortrows(population, -iQuality);

top\_q = 0;

for generations = 1 : clusterGen

if foul\_counter > 16

message = 'breaking on three fouls'

break;

end

% create the mutations

for index = 1 : population\_size \* 0.2

population{population\_size + 1 - index, iSeed} = permutate(population{sz, iSeed}, n, 5);

end

% create the hybrids

for index = population\_size/2 : 2 : 2

parent1 = population{population\_size/2 + 1 - index, iSeed};

parent2 = population{population\_size/2 + 2 - index, iSeed};

population{index + population\_size\*0.3, iSeed} = cross(parent1, parent2, n);

population{index + population\_size\*0.3 -1, iSeed} = cross(parent2, parent1, n);

end

% calculate for the bottom 14

inter\_map = [];

inter\_features = [];

top\_num = 0;

top\_classes = [];

for sz = population\_size\*0.3 + 1 : population\_size % number of samples

num = population(sz, iSeed);

[map, q, q3, q5, trust, features, cl, featuremap] = singletsne(new, n, labels);

population{sz, iSeed} = featuremap;

population{sz, iQuality} = q;

population{sz, iQuality3} = q3;

population{sz, iQuality5} = q5;

population{sz, iTrust} = trust;

population{sz, iFeatures} = features;

if q > top\_q

inter\_map = map;

inter\_features = features;

top\_q = q;

top\_classes = cl;

end

end

% Sort the existing population

population = sortrows(population, -iQuality);

legend('off');

% Genetic algorithm implementation

if size(inter\_map,1) > 0

try

foul\_counter = 0;

top\_num = population{1, iSeed};

top\_q = population{1, iQuality};

top\_q3 = population{1, iQuality3};

top\_q5 = population{1, iQuality5};

top\_trust = population{1, iTrust};

top\_features = binaryVectorToHex(population{sz});

map = inter\_map;

gscatter(inter\_map(:,1), inter\_map(:,2), labels);

if trial == 3 || trial == 8

modeText = mode{1};

else

modeText = sprintf('%d', mode(1));

end

legend(modeText, sprintf('Not %s', modeText));

message = sprintf('results/%s\_%s\_population\_%0.3f.xls',name, modeText, generations);

xlswrite(message, population);

filename = sprintf('results/%s\_%s\_q\_%0.3f\_q3\_%0.3f\_q5\_%0.3f\_trust\_%0.3f\_features\_%s.png',name, modeText, top\_q, top\_q3, top\_q5, top\_trust, top\_features);

title(sprintf('%s %s q %0.3f',name, modeText, top\_q), modeText, sprintf('Not %s', modeText));

saveas(h1, filename);

figfilename = sprintf('results/%s\_%s\_q\_%0.3f\_q3\_%0.3f\_q5\_%0.3f\_trust\_%0.3f\_features\_%s.fig',name, modeText, top\_q, top\_q3, top\_q5, top\_trust, top\_features);

savefig(h1,figfilename);

xlswrite(sprintf('results/%s\_%s\_q\_%0.3f\_q3\_%0.3f\_q5\_%0.3f\_trust\_%0.3f\_\_features\_%s.xls',name, modeText, top\_q, top\_q3, top\_q5, top\_trust, top\_features), map);

allKeys = keys(top\_classes);

csvClasses = [];

row = 1;

for i = 1:size(allKeys, 2)

csvClasses(1, row) = i; %allKeys{i};

vec = top\_classes(allKeys{i})

for j = 1: size(vec, 2)

csvClasses(j+1, row) = vec(j);

end

row = row + 1;

end

csvwrite(sprintf('results/%s\_q\_%0.3f\_classes.csv',name, top\_q), csvClasses);

catch ME

message = 'Failed writing the result files'

end

else

message = 'invalid map'

foul\_counter = foul\_counter +1;

end

end

## quality.m

This is the fitness function for the genetic algorithm.

function [quality classes] = quality(X, Y, neighbors)

% Quality tests the dimensionality reduction quality

%

% It takes the results and calculates a score:

% for each point if the nearest neighbor is of the same label it adds one

% and returns the ratio of those points.

% We take into account only the point we want to cluster

quality = 0;

[m,l] = size(X);

[mY,lY] = size(Y);

outOf = max(l, neighbors+1); % since we ignore the 1st one

[n,d]=knnsearch(X,X,'k',outOf);

total = 0;

if iscellstr(Y)

classes = containers.Map('KeyType','char', 'ValueType','any');

for i = 1 : mY

if ~isKey(classes,Y{i})

classes(Y{i}) = [0, 0]; % instancess, successes

end

end

else

classes = containers.Map('KeyType','uint64', 'ValueType','any');

for i = 1 : mY

if ~isKey(classes,Y(i))

classes(Y(i)) = [0, 0]; % instancess, successes

end

end

end

for i = 1 : m

if iscellstr(Y)

vec = classes(Y{i});

vec(1) = vec(1) + 1;

if Y{i} == 0 % meaning it's the mode

total = total + 1;

close = 0;

for op = 2 : outOf

if strcmp(Y{i},Y{n(i,op)})

close = close + 1;

end

end

if(close >= outOf/2)

quality = quality +1;

vec(2) = vec(2) + 1;

end

end

classes(Y{i}) = vec;

else

vec = classes(Y(i));

vec(1) = vec(1) + 1;

if Y(i) == 0

total = total + 1;

close = 0;

for op = 2 : outOf

if Y(i) == Y(n(i,op))

close = close +1;

end

end

if(close >= outOf/2)

quality = quality +1;

vec(2) = vec(2) + 1;

end

end

classes(Y(i)) = vec;

end

end

quality = quality / total; % m;

end

## trustworthiness.m

Get the measure of the clustering of the dimensionality reduction.

function trustworthiness = trustworthiness(X, XX)

% Trustworthiness tests the dimentionality reduction by checking whether

% the k nearest neighbors in original sample X are still the k nearest

% neighbors in the XX dimantionality reduced set

[n,d]=knnsearch(X,X,'k',100);

[nn,dd]=knnsearch(XX,XX,'k',100);

trustworthiness = 0;

[m,l] = size(n);

for i = 1 : m % go over all points

C = intersect(nn(i, :), n(i, :));

trustworthiness = trustworthiness + (length(C) -1)/(l-1); % a point is always closest to itself

end

trustworthiness = trustworthiness / m;

end

## cross.m

Cross breeding two individuals (feature set selections)

function cross = cross(parent1, parent2, maxFeatures)

% cross the numbers' bits

cross = 0;

for digit = 0:maxFeatures-1

if digit < maxFeatures/2

if bitand(parent1, 2^digit) > 0

cross = cross + 2^digit;

end

else

if bitand(parent2, 2^digit) > 0

cross = cross + 2^digit;

end

end

end

end

## permutate.m

Permutate a feature set.

function permutate = permutate(permutate, maxFeatures, minFeatures)

% Permutate changes the feature selection based on the progression

% The options are feature deletion, feature addition and feature

% renumbering

delta = 0;

while delta == 0

% select a permutation

action = rand;

temp = permutate;

if action < 0.33

% remove features

permutate = remove\_features(maxFeatures, minFeatures, permutate);

elseif action > 0.66

% add features

permutate = add\_features(maxFeatures, permutate);

else

% replace features

permutate = remove\_features(maxFeatures, minFeatures, permutate);

permutate = add\_features(maxFeatures, permutate);

end

delta = temp - permutate;

end

end

% add up to 3 features

function result = add\_features(maxFeatures, permutate)

for sz = 1 : size(permutate) % number of samples

permutate(sz) = permutate(sz) || rand > 0.5;

end

result = permutate;

end

% remove up to 3 features

function result = remove\_features(maxFeatures, minFeatures, permutate)

for sz = 1 : size(permutate) % number of samples

permutate(sz) = permutate(sz) && rand > 0.5;

end

result = permutate;

end

## single\_tsne.m

Execute a single t-SNE dimensionality reduction and analyze the results.

function [map, q1, q3, q5, trust, features, c1, featuremap] = singletsne(new, n, labels)

% cross the numbers' bits

brr = [0 0];

while brr(1) == 0

[data, features, featuremap] = dataForSeed(new, n);

[columns,rows] = size(data);

while columns < 3 % irrelevant

num = floor(rand \* 2^n);

[data, features, featuremap] = dataForSeed(new, n);

[columns,rows] = size(data);

end

% basic t-sne parameters

numDims = 2; pcaDims = 50; perplexity = 30; theta = .1 + 6/8; alg = 'svd';

% Run t-sne

map = fast\_tsne(data, numDims, pcaDims, perplexity, theta, alg);

brr = size(map);

end

[q1, c1] = quality(map, labels, 1);

[q3, c3] = quality(map, labels, 3);

[q5, c5] = quality(map, labels, 5);

% if there is improvement over the best result keep the result.

trust = trustworthiness(data, map);

end

## fast\_tsne.m

The entry point for the dimensionality reduction algorithm.

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function mappedX = fast\_tsne(X, no\_dims, initial\_dims, perplexity, theta, alg, max\_iter)

%FAST\_TSNE Runs the C++ implementation of Barnes-Hut t-SNE

%

% mappedX = fast\_tsne(X, no\_dims, initial\_dims, perplexity, theta, alg)

%

% Runs the C++ implementation of Barnes-Hut-SNE. The high-dimensional

% datapoints are specified in the NxD matrix X. The dimensionality of the

% datapoints is reduced to initial\_dims dimensions using PCA (default = 50)

% before t-SNE is performed. Next, t-SNE reduces the points to no\_dims

% dimensions. The perplexity of the input similarities may be specified

% through the perplexity variable (default = 30). The variable theta sets

% the trade-off parameter between speed and accuracy: theta = 0 corresponds

% to standard, slow t-SNE, while theta = 1 makes very crude approximations.

% Appropriate values for theta are between 0.1 and 0.7 (default = 0.5).

% The variable alg determines the algorithm used for PCA. The default is set

% to 'svd'. Other options are 'eig' or 'als' (see 'doc pca' for more details).

% The function returns the two-dimensional data points in mappedX.

%

% NOTE: The function is designed to run on large (N > 5000) data sets. It

% may give poor performance on very small data sets (it is better to use a

% standard t-SNE implementation on such data).

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% OF SUCH DAMAGE.

if ~exist('no\_dims', 'var') || isempty(no\_dims)

no\_dims = 2;

end

if ~exist('initial\_dims', 'var') || isempty(initial\_dims)

initial\_dims = 50;

end

if ~exist('perplexity', 'var') || isempty(perplexity)

perplexity = 30;

end

if ~exist('theta', 'var') || isempty(theta)

theta = 0.5;

end

if ~exist('alg', 'var') || isempty(alg)

alg = 'svd';

end

if ~exist('max\_iter', 'var') || isempty(max\_iter)

max\_iter=1000;

end

% % Perform the initial dimensionality reduction using PCA

% X = double(X);

% X = bsxfun(@minus, X, mean(X, 1));

% M = pca(X,'NumComponents',initial\_dims,'Algorithm',alg);

% X = X \* M;

% Normalize

X = normc(X);

try

% Run the fast diffusion SNE implementation

if ismac

%tic, system(fullfile(tsne\_path,'./windows/mac\_bh\_tsne')); toc

mappedX = tsne(X,'Algorithm','barneshut');

else

tsne\_path = which('fast\_tsne');

tsne\_path = fileparts(tsne\_path);

write\_data(X, no\_dims, theta, perplexity, max\_iter);

tic, system(fullfile(tsne\_path,'./windows/windows\_bh\_tsne')); toc

[mappedX, landmarks, costs] = read\_data;

landmarks = landmarks + 1; % correct for Matlab indexing

delete('data.dat');

delete('result.dat');

end

catch ME

disp('Failed on t-sne');

mappedX = [];

end

end

% Writes the datafile for the fast t-SNE implementation

function write\_data(X, no\_dims, theta, perplexity, max\_iter)

[n, d] = size(X);

h = fopen('data.dat', 'wb');

fwrite(h, n, 'integer\*4');

fwrite(h, d, 'integer\*4');

fwrite(h, theta, 'double');

fwrite(h, perplexity, 'double');

fwrite(h, no\_dims, 'integer\*4');

fwrite(h, max\_iter, 'integer\*4');

fwrite(h, X', 'double');

fclose(h);

end

% Reads the result file from the fast t-SNE implementation

function [X, landmarks, costs] = read\_data

h = fopen('result.dat', 'rb');

n = fread(h, 1, 'integer\*4');

d = fread(h, 1, 'integer\*4');

X = fread(h, n \* d, 'double');

landmarks = fread(h, n, 'integer\*4');

costs = fread(h, n, 'double'); % this vector contains only zeros

X = reshape(X, [d n])';

fclose(h);

end

Dfgh

Dfgh

Dfgh

Dfh

Df

Hd

Fghd

Fgh

dfg